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(54) PEPTIDE, BRONCHODILATOR AND BLOOD FLOW AMELIORANT

(57) Peptides represented by the following general formula (1) or pharmaceutically acceptable salts thereof, and a bronchodilator or a blood flow ameliorant containing the same as the active ingredient: H-His-Ser-Asp-A-B-Phe-Thr-Asp-C-Tyr-D-Arg-E-Arg-F-Gln-G-Ala-Val-I-J-Tyr-Leu-K-L-M-Leu-N, wherein A represents Ala or Gly; B represents Ile or Val; C represents Asn or Ser; D represents Thr or Ser; E represents Leu or Tyr; F, I and J represent each Lys or Arg, provided that at least one of F, I and J is Arg; G represents Met, Leu or Nle; K represents Asn or Ala; L represents Ser or Ala; M represents Ile or Val; and N represents -NH₂ or Asn-NH₂, excluding the case where A is Ala, B is Val, C is Asn, D is Thr, E is Leu, K is Asn, L is Ser, M is Ile and N is Asn-NH₂ at the same time.

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Description

FIELD OF THE INVENTION

The present invention relates to a novel peptide, a bronchus-expanding agent and a blood-flow-improving agent.

BACKGROUND OF THE INVENTION

VIP (vasoactive intestinal peptide) is one kind of physiologically active peptide called brain-gut peptide, the action of which is to promote blood flow and depress blood pressure. VIP was extracted from the intestine of a pig in 1970 and it consists of 28 amino acid residues (S. I. Said, V. Mutt Science, 169, 1217 (1970)).

PACAP (pituitary adenylyl cyclase activating polypeptide) was isolated from the hypothalamus of a sheep in 1989 in a bioassay system using as an indicator the activation of adenylyl cyclase from cultured pituitary cells, and it is a structurally determined peptide consisting of 38 amino acid residues (A. Miyata, A. Arimura et al., Biochem. Biophys. Res. Commun., 164, 567 (1989)). 27 amino acid residues from the N-terminal side of PACAP has the PACAP activity and the amino acid sequence of said 27 amino acids has a considerably similar structure to that of VIP.

VIP and PACAP are similar in amino acid sequence to secretin, glucagon etc. and are thus classified as peptides belonging to the glucagon family. By virtue of their strong action on vessel expansion and blood-flow promotion, VIP and PACAP are expected for use in the treatment of an ulcer, frostbite, bed sore, impotence etc. and in the restoration and growth of hair. In addition, VIP and PACAP have considerably strong relaxation action on the bronchial smooth muscle in the respiratory system, thus alleviating the smooth muscle contraction induced by stimulators such as acetylcholine, histamine, serotonin etc. Such relaxation action of VIP and PACAP differ from the relaxation action caused via adrenalin β_2 receptor by general bronchial expansion, so VIP and PACAP are expected to have an treatment effect on asthmatic spasm difficult to cure on which a stimulator of β_2 receptor does not effectively act.

Besides VIP and PACAP, a peptide consisting of 28 to 31 amino acid residues with bronchus expanding action, blood-pressure reducing action and hair restoring action is known (see Japanese Patent Application Laid-Open Publication Nos. 246,595/87, 83,012/89 and 297,498/92).

It is however desired to develop a bronchus-expanding agent and a blood-flow-improving agent superior to VIP, PACAP and other peptides in the durability of bronchus-expanding action and blood-flow increasing action.

DISCLOSURE OF THE INVENTION

The object of the present invention is to provide a novel peptide excellent in the durability of bronchus-expanding action and further excellent in blood-flow increasing action, a bronchus-expanding agent and a blood-flow-improving agent comprising said peptide as an active ingredient.

The present invention provides a peptide represented by the general formula (1):

H-His-Ser-Asp-A-B-Phe-Thr-Asp-C-Tyr-D-Arg-E-Arg-F-Gln-G-Ala-Val-I-J-Tyr-Leu-K-L-M-Leu-N (1)

wherein A is Ala or Gly; B is Ile or Val; C is Asn or Ser; D is Thr or Ser; E is Leu or Tyr; each of F, I and J is Lys or Arg, and at least one of F, I and J is Arg; G is Met, Leu or nLeu; K is Asn or Ala; L is Ser or Ala; M is Ile or Val; and N is -NH₂ or Asn-NH₂, excluding the combination where A is Ala, B

is Val, C is Asn, D is Thr, E is Leu, K is Asn, L is Ser, M is Ile, and N is Asn-NH₂, or the general formula (2):

H-His-Ser-Asp-A-B-Phe-Thr-Asp-C-Tyr-D-Arg-E-Arg-F-Gln-G-Ala-Val-I-J-Tyr-Leu-K-L-M-Leu-P-Gly-Q-R (2)

wherein A, B, C, D, E, F, G, I, J, K, L and M possess the same meanings defined as above; at least one of F, I and J is Arg; P is Asn or a chemical bond; Q is Lys, Arg, Lys-Arg, Arg-Arg or a chemical bond; and R is -OH or -NH₂, or pharmaceutically acceptable salts thereof.

Further, the present invention provides a bronchus-expanding agent comprising said peptide or a pharmaceutically acceptable salt thereof as an active ingredient.

Further, the present invention provides a blood-flow-improving agent comprising said peptide or a pharmaceutically acceptable salt thereof as an active ingredient.

Further, the present invention provides a pharmaceutical composition comprising said peptide or a pharmaceutically acceptable salt thereof and a pharmaceutically acceptable carrier.

Further, the present invention provides a method of expanding a bronchus, which comprises administering an effective amount of said peptide or a pharmaceutically acceptable salt thereof into humans.

Further, the present invention provides a method of improving a blood flow, which comprises administering an effective amount of said peptide or a pharmaceutically acceptable salt thereof into humans.

Further, the present invention provides use of said peptide or a pharmaceutically acceptable salt thereof for expanding a bronchus.

Further, the present invention provides use of said peptide or pharmaceutically acceptable salt thereof for improving a blood flow.

The peptide of the invention or a salt thereof is excellent in the durability of bronchus-expanding action resulting from the relaxation action of smooth muscle. Hence, the peptide of the invention and a salt thereof is useful as a bronchus-expanding agent.

In addition, the peptide of the invention or a salt thereof is excellent in blood-flow-increasing action and in its durability. Therefore, the peptide of the invention or a salt thereof is also useful as a blood-flow-improving agent and it can exert an improvement effect on a blood flow at a low dosage.

The abbreviations of amino acids, peptides, protective groups, solvents etc., in the present specification follow those according to International Union of Pure and Applied Chemistry (IUPAC) or International Union of Biochemistry (IUB) or conventional abbreviations in this field. Examples are shown below. If there can be optical isomers for amino acids etc., L-isomers are meant unless otherwise specified.

His: histidine residue	Ser: serine residue	Asp: aspartic acid residue
Ala: alanine residue	Val: valine residue	Phe: phenylalanine residue
Thr: threonine residue	Tyr: tyrosine residue	Asn: asparagine residue
Leu: leucine residue	Arg: arginine residue	Lys: lysine residue
Gln: glutamine residue	Met: methionine residue	Ile: isoleucine residue
Gly: glycine residue	Nle: norleucine residue	
Boc: t-butyloxycarbonyl group	Aoc: t-amyloxycarbonyl group	
Bzl: benzyl group	Z: benzyloxycarbonyl group	
Tos: p-toluenesulfonyl group	OBu: t-butylester	
OMe: methylester	OBz: benzylester	
ONP: p-nitrophenylester	Bom: benzyloxymethyl group	
TFA: trifluoroacetic acid	THF: tetrahydrofuran	
DCM: dichloromethane	DMF: dimethylformamide	
DCC: dicyclohexylcarbodiimide		
WSC: N-ethyl-N'-dimethylaminopropyl-carbodimide		
OSu: N-hydroxysuccinimide ester		
HOSu: N-hydroxysuccinimide		
HOBt: 1-hydroxybenzotriazole		
DIEA: diisopropylethylamine.		

The peptides represented by the above formula (1) or (2) can be produced by conventional means for synthesis of known peptides. For example, these peptides can be produced in e.g. an azide method, acid chloride method, acid anhydride method, mixed acid anhydrides method, DCC method, active ester method (p-nitrophenylester method, N-hydroxysuccinic acid imide ester method, cyanomethylester method etc.), a method using Woodward reagent K, carbodiimide method, oxidation-reduction method, DCC-additive (HONB, HOBt, HOSu) method etc. according to methods described in e.g. "The Peptides" Vol. 1 (1966) [Schreder and Luhke: Academic Press, New York, U.S.A.] or "Peptido Gosei" (Peptide Synthesis) [Izumiya et al., Maruzen K.K. (1975), Japan]. These methods can be applied to both solid-phase synthesis and liquid-phase synthesis. According to the above-described methods for synthesis of conventional polypeptides, a target peptide can be produced in a stepwise elongation method where each of the amino acids in said peptide is linked sequentially to the C-terminal amino acid or in a fragment condensation method where partial fragments in said peptide are synthesized and then coupled to one another.

For example, the solid-phase synthesis in the stepwise elongation method can be carried out in accordance with the method of Merrifield, R.B. (Solid Phase Peptide Synthesis, J. Amer. Chem. Soc., 85, 2149-2159 (1963)) as follows: The C-terminal amino acid of a target peptide (that is, in the present invention, the amino acid sequence of the above general formula (1) or (2)) is protected on its amino group and then coupled to insoluble resin having functional groups capable of binding to carboxyl groups. The amino group is then deprotected, and to this free amino group is bound the next amino acid with its amino group protected. These steps are repeated many times, thus elongating the amino acid chain until the histidine residue at the 1-position of the formula (1) or (2). The peptide thus obtained is then cleaved off from the resin.

The above method requires the protection (that is, linking a protective group to the amino group) and the deprotection (that is, releasing the protective group from the amino group) for the amino group participating in peptide bonding between amino acids as well as the activation of the carboxyl group participating in peptide bonding between amino acids. If necessary, a protective group is further linked to a functional group in a side chain of the amino acid.

The protective group used in protecting the amino group includes those conventionally used. Examples are benzyloxycarbonyl (Z), t-butyloxycarbonyl (Boc), t-amyloxycarbonyl (Aoc), isobornyloxycarbonyl, p-methoxybenzyloxycarbonyl, 2-chlorobenzyloxycarbonyl, adamantyloxycarbonyl, trifluoroacetyl, phthaloyl, formyl, o-nitrophenylsulphenyl, and diphenylphosphinothiyl.

The protective group used in protecting the carboxyl group includes those conventionally used. Examples are alkyl

esters such as methyl ester, ethyl ester, propyl ester, butyl ester and tert-butyl ester, benzyl ester, p-nitrobenzyl ester, methylbenzyl ester, p-chlorobenzyl ester, benzhydryl ester, benzyloxycarbonylhydrazide, tert-butyloxycarbonylhydrazide and tritylhydrazide.

The activation of the carboxyl group participating in peptide bonding can be carried out in any of the conventionally known methods described above, and the reagents etc. used in the activation can be selected from known ones. The activation of the carboxyl group involves reacting various reagents with said carboxyl group to form e.g. their corresponding acid chlorides, acid anhydrides or mixed acid anhydrides, asides, and active esters (for example, esters such as pentachlorophenol, p-nitrophenol, N-hydroxysuccinimide, N-hydroxybenztriazole, N-hydroxy-5-norbornen-2,3-dicarboxyimide etc.).

For the amino acid with a functional group in its side chain, the functional group is preferably protected during the reaction of forming a peptide bond. In particular, His, Tyr, Thr, Lys, Asp, Arg and Ser are preferably protected at the functional group in their side chain. The protection of the functional group is carried out by linking the protective group to it in a conventional method. After the completion of peptide synthesis, the protective group is released.

The protective group used in protecting the imino group in His includes e.g. benzyloxymethyl (Bom), tosyl (Tos), benzyl (Bzl), benzyloxycarbonyl (Z) and trityl.

The hydroxyl group in Ser and Thr can be protected by esterification or etherification, but its protection is not necessarily required. The protective group to be introduced by esterification includes e.g. lower alkanoyl groups such as acetyl etc., aroyl groups such as benzoyl etc., and carbonic acid-derived groups such as benzyloxycarbonyl, ethyloxycarbonyl etc. Preferable examples of protective groups introduced by etherification are benzyl (Bzl), tetrahydropyranyl and tert-butyl.

The protective group used in protecting the hydroxyl group in Tyr includes e.g. benzyl (Bzl), bromobenzyloxycarbonyl (BrZ), dichlorobenzyl ($\text{Cl}_2\text{-Bzl}$), benzyloxycarbonyl (Z), acetyl, and tosyl (Tos).

The protective group used in protecting the amino group in Lys includes e.g. benzyloxycarbonyl (Z), chlorobenzyloxycarbonyl (Cl-Z), dichlorobenzyl ($\text{Cl}_2\text{-Bzl}$), t-butyloxycarbonyl (Boc), and tosyl (Tos).

The protective group used in protecting the guanidino group in Arg includes e.g. tosyl (Tos), nitro, benzyloxycarbonyl (Z), and t-amylloxycarbonyl (Aoc).

The protection of the carboxyl group in Asp is carried out by esterification with e.g. benzyl alcohol, methanol, ethanol or tert-butanol.

The reaction of forming a peptide bond can sometimes be carried out in the presence of a condensation reagent e.g. a carbodiimide reagent such as dicyclohexylcarbodiimide (DCC), carbodiimidazole etc., or tetraethyl pyrophosphate, benzotriazole-N-hydroxytrisdimethylaminophosphonium hexafluorophosphate (Bop reagent).

The insoluble resin used in the above solid-phase synthesis may be any resin having functional groups capable of binding to carboxyl groups. Examples are benzhydrylamine resin (BHA resin), chloromethyl resin, oxymethyl resin, aminomethyl resin, p-methylbenzhydrylamine resin (MBHA resin), 4-aminomethylphenoxymethyl resin, 4-hydroxymethylphenoxymethyl resin, 4-oxymethylphenylacetamidemethyl resin (PAM resin) etc. The linking of the amino acid to the resin and cleavage of the synthesized peptide from the resin can be carried out in any of the conventionally known methods.

The solvent used in the above solid-phase synthesis may be any solvents known to be usable in forming a peptide bond, and examples are water-free or water-containing dimethylformamide (DMF), dimethyl sulfoxide (DMSO), pyridine, chloroform, dioxane, dichloromethane (DCM), tetrahydrofuran (THF), ethyl acetate, N-methylpyrrolidone, and hexamethyl phosphate triamide (HMPA). These solvents can be used singly or in combination.

The peptide thus prepared can be desalted and purified in a usual manner. For example, it can be desalted and purified by ion-exchange chromatography on DEAE-cellulose etc., partition chromatography on Sephadex LH-20, Sephadex G-25 etc., normal phase chromatography on silica gel etc., reverse phase chromatography on ODS-silica gel etc., or in high performance liquid chromatography (HPLC), etc.

The pharmaceutically acceptable salts of the peptides represented by the above formula (1) or (2) include e.g. acetate, hydrochloride, phosphate etc.

The peptide of the invention and salts thereof are useful as a bronchus-expanding agent by virtue of their bronchus-expanding action. Hence, the bronchus-expanding agent containing the peptide of the invention or a salt thereof as an active ingredient is useful in suppression and improvement of asthma etc.

As the bronchus-expanding agent, the peptide of the invention or a salt thereof can be administered as such, but usually it is mixed in a usual manner with various pharmaceutically acceptable carriers to be formed into preparations in the form of liquid, gel or solid, for oral or parenteral purposes. The administration method includes inhalation (e.g. with an aerosol agent), injection, application etc.

The dosage of the present peptide or a salt thereof when used as a bronchus-expanding agent is preferably in the range of about 1 ng/kg to 1 mg/kg (body weight)/day/person, although the dosage is determined depending on the purpose for use, symptom, the age and body weight etc. of a patient, method of administration, etc.

Further, the peptide of the invention or a salt thereof is excellent in blood-flow increasing action which is superior in durability. Hence, a blood-flow improving agent comprising the peptide of the invention or a salt thereof is effective for

treatment of an ulcer, frostbite, bed sore, impotence etc. as well as for restoration and growth of hair.

The peptide of the invention or a salt thereof when used as a blood-flow improving agent is mixed with a conventional pharmaceutically acceptable carrier to be formed into pharmaceutical preparations, preferably in the form of an injection.

For preparation of such an injection, the resulting preparation is preferably sterilized and isotonic relative to blood. Any conventionally used diluents can be used for manufacture of a preparation in the form of an injection. Example of diluents are water, physiological saline etc. In this case, a sufficient amount of common salt or glycerin to prepare an isotonic solution can be contained in a preparation in the form of an injection. This preparation can contain a conventional buffer, soothing agent, preservative etc. and if necessary a coloring agent, preservative, flavoring agent, sweetener, etc., as well as other pharmaceuticals. The above active ingredient for injection purposes can be dissolved before use in distilled water. The method of administering the resulting pharmaceutical preparation is varied depending on the form of the preparation. For example, the preparation in the form of an injection is intravenously administered singly or after being mixed with conventional complements such as amino acids etc.

The present peptide or a salt thereof when used as a blood-flow-improving agent is administered preferably in 2 to 4 portions daily in the range of about 1 ng/kg to 1 mg/kg (body weight)/day/person, although the dosage is determined depending on the purpose for use, symptom conditions, etc.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a graph showing a change with time in the degree of relaxation of bronchial smooth muscle in the case of administration of PACAP.

FIG. 2 is a graph showing a change with time in the degree of relaxation of bronchial smooth muscle in the case of administration of Peptide 5 shown in SEQ ID NO: 5.

FIG. 3 is a graph showing a change with time in the degree of relaxation of bronchial smooth muscle in the case of administration of VIP.

FIG. 4 is a graph showing a change with time in the degree of relaxation of bronchial smooth muscle in the case of administration of Peptide 11 shown in SEQ ID NO: 11.

BEST EMBODIMENTS FOR CARRYING OUT THE INVENTION

Hereinafter, the present invention is described in detail by reference to Examples, which however are not intended to limit the scope of the present invention.

In the following examples, the identification of the resulting purified peptide was carried out by measurement of retention time in high performance liquid chromatography (HPLC), measurement of degree of optical rotation, and amino acid analysis, as shown below.

- High performance liquid chromatography (HPLC)

For analysis in high performance liquid chromatography, LC-Module-1 (Waters Ltd. Japan) was used.

(Analysis conditions in HPLC)

Column : TSK Gel ODS-120T (4.6 × 250 mm)

Solvent: 0.1 % TFA-acetonitrile (linear gradient of from 20 % to 50 % acetonitrile for 30 min).

Flow rate: 1 ml/min.

Detection wavelength: 220 nm.

- Degree of optical rotation

For measurement of degree of optical rotation, DIC-370 (Nippon Bunko Kogyo K.K., Japan) was used.

(Conditions for measurement of degree of optical rotation)

Light source: Na lamp, 589 nm.

Temperature: 20°C.

Layer length: 100 mm.

Concentration: 1 % (0.1 M in acetic acid).

- Amino acid analysis:

The amino acid analysis was carried out by hydrolyzing a peptide at 110 °C for 20 hours in 6 N HCl containing 0.1 % phenol and analyzing the hydrolysate in Hitachi Amino Acid Analyzer L-8500 (Hitachi, Ltd., Japan).

Example 1

Production of peptide 1

5 The solid-phase synthesizer used was Peptide Synthesizer 9600 (Milligen Bioresearch). Peptide 1 shown in SEQ ID NO:1 whose C-terminal group was amidated was produced by the following solid-phase synthesis and purification.

First, 694 mg MBHA resin (0.72 mmol/g amino group, manufactured by Peptide Kenkyuzyo K.K., Japan) was introduced into a reactor for peptide solid-phase synthesis and treated with 8 ml DCM (4 times, 1 minute each), 8 ml DCM containing 60 % TFA (20 minutes), 4 ml DCM (3 times, 15 seconds each), 3 ml DMF solution containing 1 ml DIEA (2
10 times, 1 minute each), and 8 ml DMF (6 times, 40 seconds each) in this order with stirring in an argon stream. After each treatment, filtration was carried out.

Separately, 2 mmol amino-group-protected amino acid Boc-Leu-OH corresponding to the amino acid residue at the 27-position in SEQ ID NO:1 was dissolved in 4 ml DCM. This solution was introduced into a vessel for amino acid activation, and after 3 ml DCC (0.5 M in DCM) and 4 ml HOBt (0.5 M in DCM) were added to it, and they were reacted for
15 30 minutes. Thereafter, the reaction solution was filtered off and the filtrate was transferred to a concentration vessel. 3 ml DMF was added to it and the DCM was distilled off in an argon stream. Then, 3 ml DMF was added to the concentrate, and it was transferred to the above reactor for peptide solid-phase synthesis and reacted for 30 minutes. Then, the reaction mixture was washed with 8 ml DCM (6 times, 20 seconds each). Separately, 2 mmol Boc-Leu-OH was dissolved in 4 ml DCM and the solution was introduced into the reactor for amino acid activation and the same procedure
20 as described above was repeated (double-coupling) and filtered. Boc-Leu-MBHA resin was thus obtained.

Then, the Boc-Leu-MBHA resin was washed with 8 ml DCM (4 times, 1 minute each) and filtered. This resin was treated with 8 ml DCM containing 60 % TFA (20 minutes), 4 ml DCM (3 times, 15 seconds each), 3 ml DMF solution containing 1 ml DIEA (2 times, 1 minute each), and 8 ml DMF (6 times, 40 seconds each) in this order with stirring in an argon stream. After each treatment, filtration was carried out.

Separately, 2 mmol amino-group-protected amino acid Boc-Val-OH corresponding to the amino acid residue at the 26-position in SEQ ID NO:1 was dissolved in 4 ml DCM and introduced into a vessel for amino acid activation, and after 1.5 ml DCC (0.5 M in DCM) was added to it, and they were reacted for 7 minutes. Thereafter, the reaction solution was filtered off and the filtrate was transferred to a concentration vessel. 3 ml DMF was added to it and the DCM was distilled off in an argon stream. Then, 3 ml DMF was added to the concentrate, and it was transferred to the above reactor for
30 peptide solid-phase synthesis and reaction was carried out for 30 minutes. Then, the reaction mixture was washed with 8 ml DCM (6 times, 20 seconds each) and filtered. Boc-Val-Leu-MBHA resin was thus obtained.

Then, the following amino-group-protected amino acids, corresponding to the 25th to 1st amino acids in SEQ ID NO:1 respectively, were coupled successively to it.

Amino Acid No.	Protected Amino Acid	Amount (mmol)
25	Boc-Ala-OH	2
24	Boc-Ala-OH	2 × 2
23	Boc-Leu-OH	2
22	Boc-Tyr(Bzl)-OH	2
21	Boc-Arg(Tos)-OH	2 × 2
20	Boc-Arg(Tos)-OH	2 × 2
19	Boc-Val-OH	2
18	Boc-Ala-OH	2
17	Boc-Leu-OH	2
16	Boc-Gln-OH	2 × 2
15	Boc-Arg(Tos)-OH	2 × 2
14	Boc-Arg(Tos)-OH	2 × 2
13	Boc-Tyr(Bzl)-OH	2
12	Boc-Arg(Tos)-OH	2 × 2
11	Boc-Ser(Bzl)-OH	2
10	Boc-Tyr(Bzl)-OH	2
9	Boc-Ser(Bzl)-OH	2
8	Boc-Asp(OBz)-OH	2
7	Boc-Thr(Bzl)-OH	2
6	Boc-Phe-OH	2
5	Boc-Ile-OH	2
4	Boc-Gly-OH	2
3	Boc-Asp(OBz)-OH	2
2	Boc-Ser(Bzl)-OH	2
1	Boc-His(Bom)-OH	2 × 2

In the above solid-phase synthesis, double coupling was carried out in the case of Asn, Arg, Gln and His.

2.76 g of the following protected peptide-HBHA resin was thus obtained:

Boc-His(Bom)-Ser(Bzl)-Asp(OBz)-Gly-Ile-Phe-Thr(Bzl)-Asp(OBz)-Ser(Bzl)-Tyr(Bzl)-Ser(Bzl)-Arg(Tos)-Tyr(Bzl)-Arg(Tos)-Arg(Tos)-Gln-Leu-Ala-Val-Arg(Tos)-Arg(Tos)-Tyr(Bzl)-Leu-Ala-Ala-Val-Leu-MBHA resin.

5 ml anisole was added to 2.76 g of the above protected peptide-MBHA resin, and 25 ml anhydrous hydrogen fluoride was added to it and the mixture was reacted with stirring at 0°C for 1 hour. Thereafter, the anhydrous hydrogen fluoride was distilled off under reduced pressure. The residue was washed with ether and then the peptide was extracted from it with 100 ml of 0.1 M acetic acid.

The extract was stirred for 15 minutes together with 20 ml anion-exchange resin Amberlite IR-410 and the insoluble resin was removed by filtration. The suspension thus obtained was filtered through 0.22 μ Millipore filter and the filtrate was lyophilized. 823 mg white powder was thus obtained. Then, this sample was applied to a column of CM-cellulose (2.5 × 30 cm) and eluted with a linear gradient of from 0.05 M to 0.5 M AcONH₄ (pH 7.0), and fraction Nos. 69 to 82 (10 ml/fraction) were combined and gave 382 mg of the desired peptide in partially purified form. This peptide was further purified in preparative high performance liquid chromatography.

Column: TSK Gel ODS-120T (21.5 × 300 mm).

Solvent: 0.1 % TFA-acetonitrile (linear gradient of from 20 % to 40 % acetonitrile).

Flow rate: 10 ml/min.

The eluate corresponding to a peak of peptide 1 as the desired substance was lyophilized. The peptide in purified form was obtained in said HPLC in an amount of 63 mg per 100 mg of the partially purified peptide. The purified peptide 1 was determined for retention time in HPLC and for degree of optical rotation, and its amino acids were analyzed. The results are shown below:

- Retention time in HPLC: 27.2 min.
- $[\alpha]_D$: -55.8°
- Amino acid analysis:
Asp(2) 2.17, Thr(1) 0.98, Ser(3) 2.45, Glu(1) 1.09, Gly(1) 1.03, Ala(3) 3.18, Val(2) 2.14, Ile(1) 0.96, Leu(3) 3.07, Tyr(3) 2.73, Phe(1) 0.92, His(1) 1.12, Arg(5) 5.16.

Example 2

Peptide 2 shown in SEQ ID NO:2 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 73 mg
- Retention time in HPLC: 27.6 min.
- $[\alpha]_D$: -53.2°
- Amino acid analysis:
Asp(2) 2.10, Thr(1) 0.97, Ser(3) 2.46, Glu(1) 0.97, Gly(2) 1.97, Ala(3) 3.15, Val(2) 2.13, Ile(1) 1.00, Leu(3) 3.08, Tyr(3) 2.75, Phe(1) 0.98, His(1) 0.99, Arg(5) 4.54.

Example 3

Peptide 3 shown in SEQ ID NO:3 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 88 mg
- Retention time in HPLC: 27.0 min.
- $[\alpha]_D$: -52.8°
- Amino acid analysis:
Asp(2) 2.11, Thr(1) 0.95, Ser(3) 2.56, Glu(1) 0.97, Gly(2) 2.00, Ala(3) 3.15, Val(2) 2.10, Ile(1) 1.00, Leu(3) 3.08, Tyr(3) 2.75, Phe(1) 0.98, His(1) 1.03, Arg(5) 5.14, Lys(1) 1.01.

Example 4

Peptide 4 shown in SEQ ID NO:4 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 84 mg
- Retention time in HPLC: 25.1 min.
- $[\alpha]_D$: -54.3°
- Amino acid analysis:
Asp(2) 2.09, Thr(1) 0.96, Ser(3) 2.44, Glu(1) 0.97, Gly(2) 2.01, Ala(3) 3.11, Val(2) 2.03, Ile(1) 1.02, Leu(3) 3.08, Tyr(3) 2.74, Phe(1) 0.97, His(1) 1.05, Arg(6) 6.10

Example 5

Peptide 5 shown in SEQ ID NO:5 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 76 mg

- Retention time in HPLC: 24.4 min.
- $[\alpha]_D$: -52.2°
- Amino acid analysis:
Asp(2) 2.11, Thr(1) 0.96, Ser(3) 2.47, Glu(1) 1.01, Gly(2) 2.00, Ala(3) 3.03, Val(2) 2.03, Ile(1) 1.02, Leu(3) 3.03,
Tyr(3) 2.70, Phe(1) 0.97, His(1) 1.07, Arg(6) 6.15, Lys(1) 1.02.

Example 6

Peptide 6 shown in SEQ ID NO:6 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 85 mg
- Retention time in HPLC: 23.6 min.
- $[\alpha]_D$: -53.7°
- Amino acid analysis:
Asp(2) 2.09, Thr(1) 0.94, Ser(3) 2.51, Glu(1) 1.05, Gly(2) 2.00, Ala(3) 3.03, Val(2) 2.01, Ile(1) 1.02, Leu(3) 3.03,
Tyr(3) 2.77, Phe(1) 0.97, His(1) 1.07, Arg(7) 7.11.

Example 7

Peptide 7 shown in SEQ ID NO:7 was obtained in purified form in the same manner as in Example 1 except that 704 mg of Boc-Arg(Tos)-PAM resin (0.68 mmol/g amino group, produced by Peptide Kenkyuzyo K.K.), i.e. a resin having the C-terminal amino acid residue of the target peptide, was used. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 84 mg
- Retention time in HPLC: 23.8 min.
- $[\alpha]_D$: -56.7°
- Amino acid analysis:
Asp(2) 2.12, Thr(1) 0.96, Ser(3) 2.47, Glu(1) 1.01, Gly(2) 2.00, Ala(3) 3.03, Val(2) 2.03, Ile(1) 1.02, Leu(3) 3.03,
Tyr(3) 2.70, Phe(1) 0.97, His(1) 1.03, Arg(6) 6.10, Lys(1) 1.02.

Example 8

Peptide 8 shown in SEQ ID NO:8 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 68 mg
- Retention time in HPLC: 26.9 min.
- $[\alpha]_D$: -58.7°
- Amino acid analysis:
Asp(5) 5.11, Thr(2) 1.72, Ser(2) 1.61, Glu(1) 1.10, Ala(2) 2.00, Val(2) 2.00, Ile(1) 1.02, Leu(4) 4.15, Tyr(2) 1.86,
Phe(1) 0.99, His(1) 0.95, Arg(5) 4.75.

Example 9

Peptide 9 shown in SEQ ID NO:9 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 76 mg
- Retention time in HPLC: 25.4 min.
- $[\alpha]_D$: -52.9°
- Amino acid analysis:
Asp(5) 5.11, Thr(2) 1.83, Ser(2) 1.49, Glu(1) 1.09, Gly(1) 1.04, Ala(2) 2.01, Val(2) 2.01, Ile(1) 0.96, Leu(4) 4.19,
Tyr(2) 2.03, Phe(1) 1.01, His(1) 1.07, Arg(6) 6.10, Lys(1) 1.05.

Example 10

Peptide 10 shown in SEQ ID NO:10 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 77 mg
- Retention time in HPLC: 24.9 min.
- $[\alpha]_D$: -52.5°
- Amino acid analysis:
 Asp(5) 5.12, Thr(2) 1.83, Ser(2) 1.55, Glu(1) 1.08, Gly(1) 1.03, Ala(2) 2.01, Val(2) 2.01, Ile(1) 0.96, Leu(4) 4.17, Tyr(2) 2.03, Phe(1) 1.01, His(1) 1.03, Arg(6) 6.15.

Example 11

Peptide 11 shown in SEQ ID NO:11 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 87 mg
- Retention time in HPLC: 23.5 min.
- $[\alpha]_D$: -55.5°
- Amino acid analysis:
 Asp(5) 5.11, Thr(2) 1.83, Ser(2) 1.49, Glu(1) 1.09, Gly(1) 1.04, Ala(2) 2.01, Val(2) 2.01, Ile(1) 0.96, Leu(4) 4.19, Tyr(2) 2.03, Phe(1) 1.01, His(1) 1.07, Arg(6) 6.10, Lys(1) 1.05.

Example 12

Peptide 12 shown in SEQ ID NO:12 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 78 mg
- Retention time in HPLC: 23.0 min.
- $[\alpha]_D$: -56.2°
- Amino acid analysis:
 Asp(5) 5.13, Thr(2) 1.84, Ser(2) 1.49, Glu(1) 1.08, Gly(1) 1.03, Ala(2) 2.01, Val(2) 2.01, Ile(1) 0.96, Leu(4) 4.19, Tyr(2) 2.03, Phe(1) 1.01, His(1) 1.02, Arg(7) 7.11.

Example 13

Peptide 13 shown in SEQ ID NO:13 was obtained in purified form in the same manner as in Example 1 except that 704 mg of Boc-Arg(Tos)-PAM resin, i.e. a resin having the C-terminal amino acid residue of the target peptide, was used. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 83 mg
- Retention time in HPLC: 22.8 min.
- $[\alpha]_D$: -58.6°
- Amino acid analysis:
 Asp(5) 5.15, Thr(2) 1.83, Ser(2) 1.53, Glu(1) 1.09, Gly(1) 1.01, Ala(2) 2.01, Val(2) 2.01, Ile(1) 1.02, Leu(4) 4.10, Tyr(2) 1.86, Phe(1) 1.00, His(1) 0.95, Arg(6) 6.15.

Example 14

Peptide 14 shown in SEQ ID NO:14 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 64 mg
 - Retention time in HPLC: 25.1 min.
 - $[\alpha]_D$: -52.7°
 - Amino acid analysis:
- 5 Asp(2) 2.17, Thr(1) 0.98, Ser(3) 2.55, Glu(1) 1.03, Gly(1) 1.03, Ala(3) 3.18, Met(1) 0.96, Val(2) 2.14, Ile(1) 0.96, Leu(2) 2.07, Tyr(3) 2.73, Phe(1) 1.00, His(1) 1.01, Arg(5) 5.15.

Example 15

10 Peptide 15 shown in SEQ ID NO:15 whose C-terminal carboxyl group was amidated was obtained in purified form in the same manner as in Example 1. The yield of this purified peptide is shown below. Its retention time in HPLC and its degree of optical rotation were determined, and its amino acids were analyzed. The results are shown below:

- Yield: 62 mg
 - Retention time in HPLC: 26.2 min.
 - $[\alpha]_D$: -52.5°
 - Amino acid analysis:
- 15 Asp(2) 2.11, Thr(1) 0.95, Ser(3) 2.45, Glu(1) 1.04, Gly(1) 1.03, Ala(3) 3.18, Val(2) 2.14, Ile(1) 0.96, Leu(2) 2.10, Tyr(3) 2.73, Phe(1) 0.96, His(1) 1.02, Arg(5) 5.16, nLeu(1) 1.02.

Example 16

Purified peptides 1 to 15 obtained in Examples 1 to 15 were examined for their bronchus-expanding action in the following manner and compared with that of VIP and PACAP.

25 Male guinea pigs each weighing 450 to 500 g were cut at the jugular veins to open the chests, and immediately the tracheae were immediately removed. Their tracheae were immediately placed in a Krebs solution and cut into loops and then formed into a chain by linking the cartilage parts with one another via a steel string. Then, the cartilage parts in the opposite side to the smooth muscular parts were cut such that the smooth muscular parts were attached to one another. The resulting sample was placed in an about 10-ml isothermic organ bath and perpendicularly suspended.

30 The lower part was fixed and the upper part was connected to a grass force transducer with a load of 1.4 g. A gas of 94 % oxygen and 6 % carbon dioxide was sufficiently blown into this sample while a Krebs solution containing 0.1 μ mol/l carbachol was dropped to it at a rate of 0.33 ml/min. to determine relaxation action. Because the relaxation reaction occurs depending on the content of a test sample in this drop solution, this can be used to compare the relaxation action of VIP and PACAP and that of the peptides of the invention on the bronchial smooth muscle.

35 A solution of each of VIA, PACAP and the peptides of the invention was prepared in a concentration as high as 100-fold than the final concentration so that its concentration in the organ bath could be adjusted. 30 minutes after the carbachol-containing Krebs solution was dropped, 100 μ l of the sample solution was dropped.

40 It was assumed that the degree of contraction of the smooth muscle is 0 in the absence of carbachol and the degree is 100 in the presence of carbachol. Minimum contraction degree A (i.e. maximum relaxation value) in the presence of the sample was determined, and maximum relaxation degree B was calculated as follows:

$$\text{Maximum relaxation degree B (\%)} = 100 - A$$

45 The time required to reach half of the maximum relaxation value (referred to hereinafter as "half-time T") after the addition of the sample was determined.

50 A change in the relaxation degree of the bronchial smooth muscle with time was determined using each peptide at 3 dosages of 0.3 μ M, 1 μ M, and 3 μ M respectively. The results are shown in the drawings: PACAP in FIG. 1; peptide 5 obtained in Example 5, in FIG. 2; VIP in FIG. 3; and peptide 11 obtained in Example 11, in FIG. 4. The maximum relaxation degree B at a dosage of 3 μ M is 75 % for peptide 5 and 80 % for peptide 11, and half-time T is 360 minutes or more in both the cases. From these results, the peptides of the invention, while possessing a similar relaxation effect to those of VIP and PACAP, exhibit superior durability, i.e. a longer working period than those of VIP and PACAP.

Table 1 shows maximum relaxation degree B and half-life T for each peptide at each dosage.

Table 1

	0.3 μ M		1 μ M		3 μ M	
	B (%)	T (min)	B (%)	T (min)	B (%)	T (min)
PACAP	40	60	90	60	90	100
VIP	45	130	80	150	85	240
Peptide 1 in Example 1	50	100	80	240	85	>360
Peptide 2 in Example 2	55	120	80	240	85	>360
Peptide 3 in Example 3	55	120	80	240	85	>360
Peptide 4 in Example 4	55	120	80	240	85	>360
Peptide 5 in Example 5	60	120	80	>360	75	>360
Peptide 6 in Example 6	65	150	80	>360	75	>360
Peptide 7 in Example 7	60	120	80	>360	75	>360
Peptide 8 in Example 8	50	150	80	300	80	>360
Peptide 9 in Example 9	55	180	80	300	80	>360
Peptide 10 in Example 10	55	200	80	300	80	>360
Peptide 11 in Example 11	65	300	85	>360	85	>360
Peptide 12 in Example 12	65	300	90	>360	85	>360
Peptide 13 in Example 13	65	300	90	>360	85	>360
Peptide 14 in Example 14	40	100	80	240	85	>360
Peptide 15 in Example 15	40	100	80	240	85	>360

Example 17

Purified peptides 1 to 15 obtained in Examples 1 to 15 were examined for their blood-flow increasing action and compared with those of VIP and PACAP.

Wistar male rats each weighing 220 to 250 g were anesthetized by intraperitoneal administration of pentobarbital (25 mg/kg) and fixed on their back. An arterial canula was inserted into the carotid and then connected to a blood-pressure transducer (DX-300, Nippon Koden, Japan) via a doom kit (SCK-512, Nippon Koden) filled with heparin-containing physiological saline (10 U/ml) to measure blood pressure with a strain blood-pressure amplifier (AP-601G, Nippon Koden). Further, a blood-flow measurement probe (FR-030T, Nippon Koden) was attached to the right femoral artery and then connected to an electromagnetic blood-flow meter (MFV-3200, Nippon Koden) to measure blood flow. The dosage of each peptide intraarterially administrated was 100 pmol/kg.

Table 2 shows an increase in blood flow in rats given each peptide.

Table 2

	Blood Flow (ml/min)
PACAP	16.04±5.92
VIP	10.18±3.38
Peptide 1 in Example 1	50.02±42.34
Peptide 2 in Example 2	52.63±39.58
Peptide 3 in Example 3	52.13±37.85
Peptide 4 in Example 4	51.87±37.24
Peptide 5 in Example 5	54.60±41.05
Peptide 6 in Example 6	53.71±46.46
Peptide 7 in Example 7	55.05±46.71
Peptide 8 in Example 8	53.75±37.31
Peptide 9 in Example 9	54.26±35.95
Peptide 10 in Example 10	55.86±34.69
Peptide 11 in Example 11	57.04±38.67
Peptide 12 in Example 12	57.34±41.88
Peptide 13 in Example 13	56.76±36.15
Peptide 14 in Example 14	52.92±32.12
Peptide 15 in Example 15	52.63±31.05

No particular change was observed in the blood pressure of rats given 100 pmol/kg of each of peptides 1 to 15.

Example 18

Acute Toxicity Test

Peptides 1 to 15 obtained in Examples 1 to 15 were dissolved in physiological saline and administered intravenously into mice at a dosage of 10 mg/kg. The result indicated that every mouse survived.

SEQUENCE LISTING

SEQ ID NO: 1:

LENGTH: 27 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 1:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Ala Ala Val Leu

20 25

SEQ ID NO: 2:

LENGTH: 28 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 2:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Ala Ala Val Leu Gly

20 25

SEQ ID NO: 3:

LENGTH: 29 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 3:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Ala Ala Val Leu Gly Lsy

20

25

SEQ ID NO: 4:

LENGTH: 29 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 4:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1

5

10

15

Leu Ala Val Arg Arg Tyr Leu Ala Ala Val Leu Gly Arg

20

25

SEQ ID NO: 5:

LENGTH: 30 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 5:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1

5

10

15

Leu Ala Val Arg Arg Tyr Leu Ala Ala Val Leu Gly Lys Arg

20

25

30

SEQ ID NO: 6:

LENGTH: 30 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 6:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1 5 10 15
 Leu Ala Val Arg Arg Tyr Leu Ala Ala Val Leu Gly Arg Arg
 5 20 25 30

SEQ ID NO: 7:

LENGTH: 30 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 7:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Gln

1 5 10 15
 Leu Ala Val Arg Arg Tyr Leu Ala Ala Val Leu Gly Lys Arg
 20 25 30

SEQ ID NO: 8:

LENGTH: 29 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 8:

His Ser Asp Ala Val Phe Thr Asp Asn Tyr Thr Arg Leu Arg Arg Gln

1 5 10 15
 Leu Ala Val Arg Arg Tyr Leu Asn Ser Ile Leu Asn Gly
 20 25

SEQ ID NO: 9:

LENGTH: 30 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 9:

His Ser Asp Ala Val Phe Thr Asp Asn Tyr Thr Arg Leu Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Asn Ser Ile Leu Asn Gly Lys

20 25 30

SEQ ID NO: 10:

LENGTH: 30 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 10:

His Ser Asp Ala Val Phe Thr Asp Asn Tyr Thr Arg Leu Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Asn Ser Ile Leu Asn Gly Arg

20 25 30

SEQ ID NO: 11:

LENGTH: 31 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 11:

His Ser Asp Ala Val Phe Thr Asp Asn Tyr Thr Arg Leu Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Asn Ser Ile Leu Asn Gly Lys Arg

20 25 30

SEQ ID NO: 12:

LENGTH: 31 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 12:

His Ser Asp Ala Val Phe Thr Asp Asn Tyr Thr Arg Leu Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Asn Ser Ile Leu Asn Gly Arg Arg

20 25 30

SEQ ID NO: 13:

LENGTH: 31 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 13:

His Ser Asp Ala Val Phe Thr Asp Asn Tyr Thr Arg Leu Arg Arg Gln

1 5 10 15

Leu Ala Val Arg Arg Tyr Leu Asn Ser Ile Leu Asn Gly Lys Arg

20 25 30

SEQ ID NO: 14:

LENGTH: 27 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 14:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1 5 10 15

Met Ala Val Arg Arg Tyr Leu Ala Ala Val Leu

20 25

SEQ ID NO: 15:

LENGTH: 27 amino acids

TYPE: amino acid

TOPOLOGY: linear

MOLECULE TYPE: peptide

SEQUENCE DESCRIPTION: SEQ ID NO: 15:

His Ser Asp Gly Ile Phe Thr Asp Ser Tyr Ser Arg Tyr Arg Arg Gln

1 5 10 15

Nle Ala Val Arg Arg Tyr Leu Ala Ala Val Leu

20 25

Claims

1. A peptide represented by the general formula (1):

H-His-Ser-Asp-A-B-Phe-Thr-Asp-C-Tyr-D-Arg-E-Arg-F-Gln-G-Ala-Val-I-J-Tyr-Leu-K-L-M-Leu-N (1)

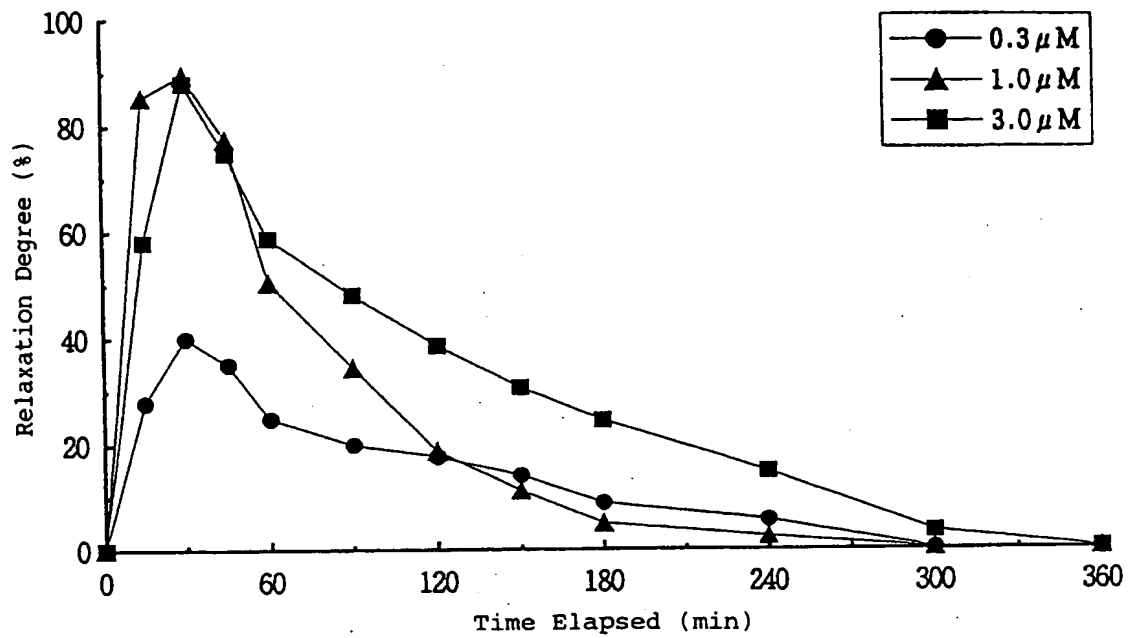
wherein A is Ala or Gly; B is Ile or Val; C is Asn or Ser; D is Thr or Ser; E is Leu or Tyr; each of F, I and J is Lys or Arg, and at least one of F, I and J is Arg; G is Met, Leu or nLeu; K is Asn or Ala; L is Ser or Ala; M is Ile or Val; and N is -NH₂ or Asn-NH₂, excluding the combination where A is Ala, B is Val, C is Asn, D is Thr, E is Leu, K is Asn, L is Ser, M is Ile, and N is Asn-NH₂, or the general formula (2):

H-His-Ser-Asp-A-B-Phe-Thr-Asp-C-Tyr-D-Arg-E-Arg-F-Gln-G-Ala-Val-I-J-Tyr-Leu-K-L-M-Leu-P-Gly-Q-R (2)

wherein A, B, C, D, E, F, G, I, J, K, L and M possess the same meanings as defined above; at least one of F, I and J is Arg; P is Asn or a chemical bond; Q is Lys, Arg, Lys-Arg, Arg-Arg or a chemical bond; and R is -OH or -NH₂, or pharmaceutically acceptable salts thereof.

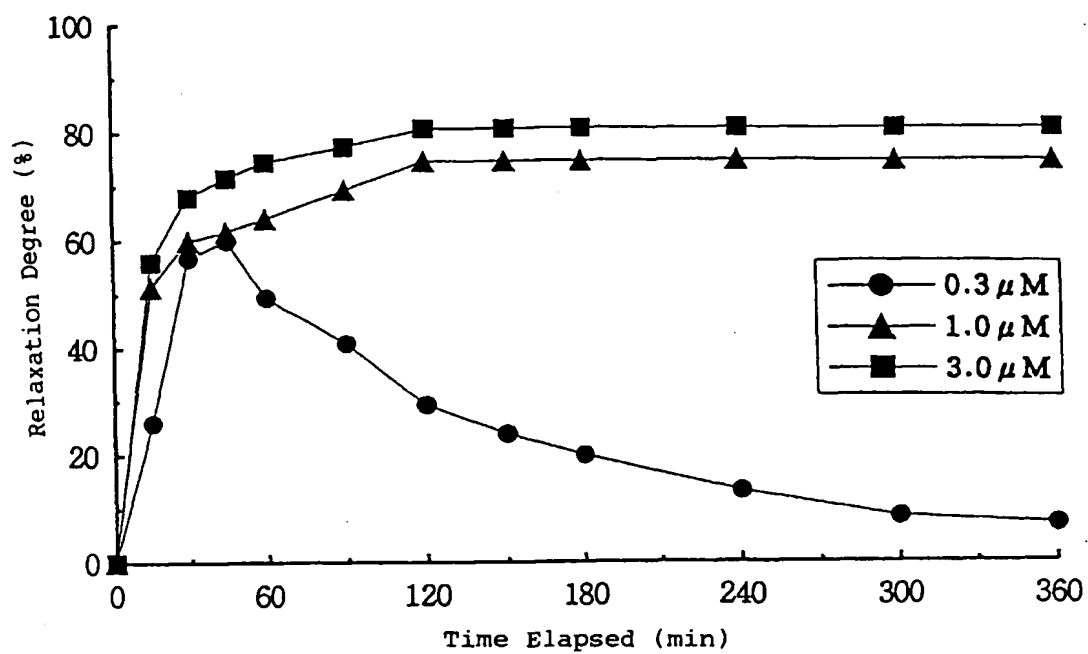
2. A bronchus-expanding agent comprising the peptide of claim 1 or a pharmaceutically acceptable salt thereof as an active ingredient.
3. A blood-flow-improving agent comprising the peptide of claim 1 or a pharmaceutically acceptable salt thereof as an active ingredient.
4. A pharmaceutical composition comprising the peptide of claim 1 or a pharmaceutically acceptable salt thereof and a pharmaceutically acceptable carrier.
5. A method of expanding a bronchus, which comprises administering an effective amount of the peptide of claim 1 or a pharmaceutically acceptable salt thereof into humans.
6. A method of improving a blood flow, which comprises administering an effective amount of the peptide of claim 1 or a pharmaceutically acceptable salt thereof into humans.
7. Use of the peptide of claim 1 or a pharmaceutically acceptable salt thereof for expanding a bronchus.
8. Use of the peptide of claim 1 or a pharmaceutically acceptable salt thereof for improving a blood flow.

FIG.1



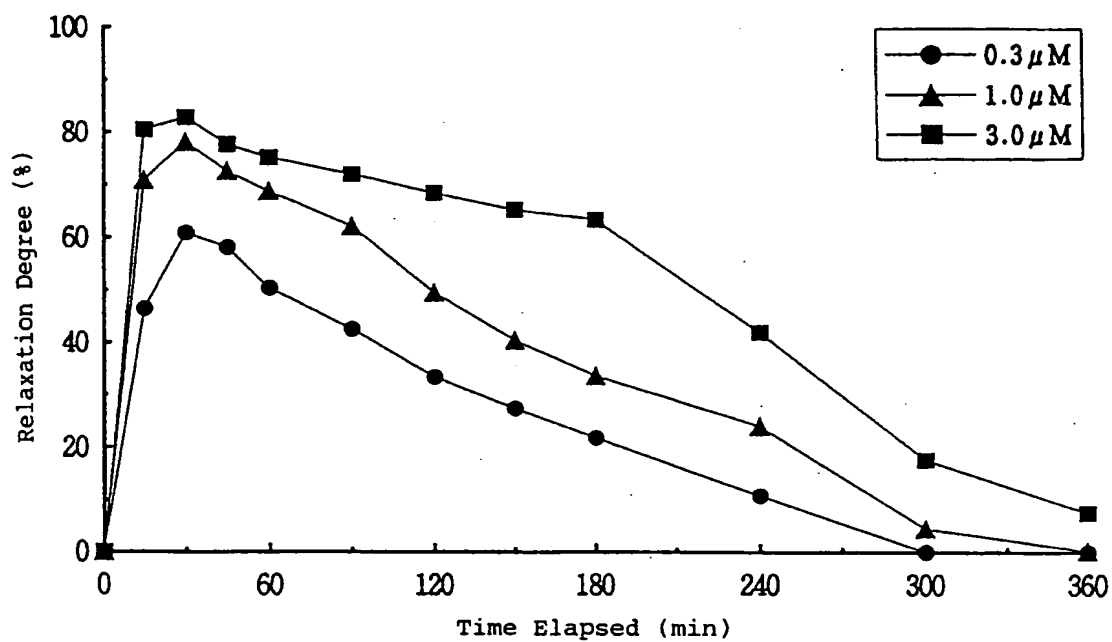
Relaxation Action of PACAP

FIG.2



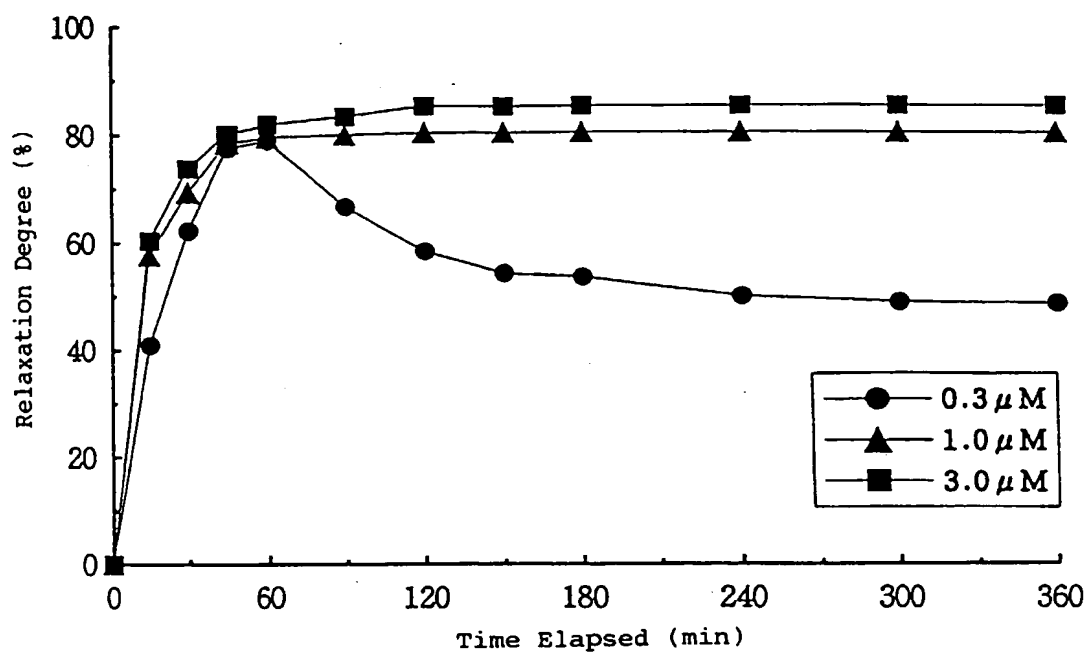
Relaxation Action of Example 5

FIG.3



Relaxation Action of VIP

FIG.4



Relaxation Action of Example 11

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP96/01543

A. CLASSIFICATION OF SUBJECT MATTER		
Int. Cl ⁶ C07K14/605, C07K14/575, A61K38/16		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols)		
Int. Cl ⁶ C07K14/605, C07K14/575, A61K38/16		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)		
CAS ONLINE		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO, 88/06598, A1 (The Salk Institute Biotechnology/Industrial Associates, Inc.), September 7, 1988 (07. 09. 88) & AU, 8814842, A & US, 4835252, A & US, 4939224, A	1 - 4
A	JP, 4-297498, A (Seikagaku Corp.), October 21, 1992 (21. 10. 92) (Family: none)	1 - 4
A	JP, 62-246595, A (Eisai Co., Ltd.), October 27, 1987 (27. 10. 87) & EP, 241926, B1 & US, 4757133, A & US, 4822774, A	1 - 4
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search August 15, 1996 (15. 08. 96)		Date of mailing of the international search report August 27, 1996 (27. 08. 96)
Name and mailing address of the ISA/ Japanese Patent Office Facsimile No.		Authorized officer Telephone No.

Form PCT/ISA/210 (second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP96/01543

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 5 - 8
because they relate to subject matter not required to be searched by this Authority, namely:
Claims 5 to 8 pertain to methods for treatment of the human or animal body by therapy and thus relate to a subject matter which this International Searching Authority is not required, under the provisions of Article 17(2)(a)(i) of the PCT and Rule 39.1(iv) of the Regulations under the PCT, to search.
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

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